# Growth and metamorphosis of *Bufo gargarizans gargarizans* larvae: effects of water volume and food diversity

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	5	Growth and metamorphosis of <i>Bufo gargarizans gargarizans</i> larvae:
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, -	14	Submitted on: 13 <sup>th</sup> December 2023; revised on: 20th June 2024; accepted on: 21st
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ź	18	Running title: Growth and metamorphosis of Bufo gargarizans gargarizans larvae
-	19	
	20	Abstract. Organisms can evolve and produce different phenotypes in respond to
4	21	local environmental conditions. Abiotic factors, such as water volume and food
2	22	diversity in breeding ponds, also play significant roles in the survival and age at
4	23	metamorphosis of amphibians. Here, we experimentally examined the plasticity of
2	24	growth rate, survival rate, larval mass, age, and body size at metamorphosis iin Bufo
	25	gargarizans gargarizans in response to varying combinations of water volume and
2	26	food diversity. The interaction between water volume and food diversity had no effect

on larval growth and development. However, food diversity had a significant impact 27 on the age at metamorphosis, with two food resources resulting in a shorter larval 28 29 period and earlier metamorphosis, particularly in larger water volumes. Our findings demonstrated that tadpoles raised in larger water volumes had larger body sizes at 30 31 metamorphosis compared to those in smaller water volumes. Additionally, tadpoles in larger water volumes exhibited higher growth rates and shorter larval period than 32 those in smaller water volumes. We suggest that the increased frequency of physical 33 encounters between tadpoles and the vessel walls in smaller volumes, akin to 34 high-density conditions, causes psychological stress due to crowding, which hinders 35 larval growth and development. 36

Keywords. *Bufo gargarizans gargarizans*, water volume, food diversity, phenotypic
plasticity, metamorphic size.

39

## INTRODUCTION

42	Organism have the ability to adapt to specific environmental conditions by
43	changing its phenotype (Pigliucci, 2001; DeWitt and Scheiner, 2004). Biotic factors
44	such as population density (Kehr et al., 2014), competition, and predation can impact
45	the composition of tadpole populations (e.g., tadpole size, developmental stage,
46	distribution, richness, and diversity). Additionally, abiotic factors like hydroperiod,
47	habitat size, or water volume can affect the survival and timing of metamorphosis in
48	amphibians (Semlitsch et al., 1996; Wilbut, 1997).
49	For organisms with complex life cycles, such as amphibians, the growth and
50	developmental rate of the larval stage can change according to different
51	environmental conditions (Richter-Boix et al., 2011). Although excessive water
52	volume can reduce the growth of certain species (Pearman, 1993; Kehr et al., 2014),
53	growth rates increase with larger absolute water volume in most cases (Pearman, 1993;
54	Montealegre-Delgado et al., 2013). Several studies have examined the potential
55	impact of abiotic factors, such as habitat space or water volume, on the density effects
56	(biotic factor) in organisms. As a result, the crowding effect caused by high
57	population density often reduces the growth and development of larvae (Smith-Gill
58	and Berven, 1979; Semlitsch and Caldwell, 1982; Relyea and Hoverman, 2003). To
59	date, four primary factors have been suggested to explain the decrease in individual
60	growth and the lengthening of the larval period. These factors include a decrease in
61	food ration (Wilbur, 1977; Hota and Dash, 1981), competition for limited food
62	resources (Brockelman, 1969; Wilbur and Collins, 1973; DeBenedictis, 1974), social

63 interactions (Gromko et al., 1973), and stress (Lynn and Edelman, 1936; John and
64 Fenster, 1975).

65 Most anuran tadpoles are herbivorous or omnivorous, but there are a few that are carnivorous and cannibalistic (Kamat, 1962; Costa and Balasubramaniam, 1965; 66 Sabnis and Kolhatkar, 1977; Sabnis and Kuthe, 1978; Sekar, 1992; Altig et al., 2007). 67 For instance, Bufo stomaticus larvae showed optimal growth when provided with a 68 diet containing a combination of animal and plant proteins. However, when these 69 larvae were only provided with goat meat, their growth rate declined, and the timing 70 of metamorphosis was delayed (Saidapur, 2001). Similarly, Bufo melanostictus larvae 71 showed comparable growth rates when fed a diet of spinach and Chironomous larvae. 72 However, their growth was stunted and metamorphosis was delayed when provided 73 with only Chironomous larvae (Sabnis and Kuthe, 1978). This suggests that 74 competition among larvae may be reduced when multiple food sources are available 75 in breeding sites, allowing individuals to utilize alternative resources (Martin and 76 77 Garnett, 2013).

Extensive studies have been conducted on the relationships between crowding and growth rates (Goetsch, 1924; Gromko et al., 1973; Steinwascher, 1978; Fixari III et al., 2017), but little attention has been paid to the impact of confining solitary tadpoles and the potential effects of water volume and food diversity on tadpole growth and development. In this study, we investigated the potential interactive effects of food diversity and water volume on the plasticity of metamorphic traits in *Bufo gargarizans gargarizans*, including the length of larval period, survival, the size at metamorphosis, and growth rate.

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#### **MATERIALS AND METHODS**

88 *Field procedures* 

*Bufo gargarizans gargarizans* exhibits sexual dimorphism and has a wide distribution in East Asia. They are explosive breeders (Wells, 2007), typically breeding between 6 to 14 days (Yu and Sharma, 2012) and tadpoles hatch after two weeks. Ponds used as breeding sites are often rich in *Spirogyra* and pondweed (e.g., *Potamogeton crispus*), which serve as a food source for tadpoles (Wei et al., 2011).

During the peak breeding period in mid-February 2018, we captured 5 mating 94 pairs of B. g. gargarizans in Shihe County (32°08'39"N, 114°02'37"E, altitude 84 m), 95 central plains of China. Each pair was then placed individually in a plastic container 96 (20 L) filled with approximately 12-15 cm of pond water until females laid their eggs. 97 Once the oviposition process was completed, we collected 50 eggs from each of the 5 98 egg masses. All eggs were placed separately into five plastic containers (5 L) filled 99 with approximately 10 cm of tap water. Tap water has been previously stored in two 100 large buckets (100 L) for several days before use. After the breeding was finished, we 101 transported all toads and the remaining eggs back to their original spawning site. 102

103

104 Laboratory procedures

We employed a randomized block design to test the effect of food diversity (two
food resources: *Potamogeton crispus* and fish food with high protein content (>45%),

lipids (>12%), algae (>12%), fiber (>4%), and ash (<10%); single food resource: P. 107 crispus), and water volume (250 mL, 500 mL, 2000 mL, 8000 mL) on the growth and 108 development of B. g. gargarizans tadpoles. Each tadpole (Gosner stage 25, absorption 109 of external gills and formed spiracle, Gosner, 1960) was placed individually in a 110 series of numbered opaque tanks. A total of 120 tanks were used, with the tank 111 diameters ranging from approximately 9.49 cm to 26.99 cm, representing the four 112 different water volumes. Water temperature was recorded using a mercury 113 thermometer. 114

The experiment was conducted using 120 tanks, with each of the eight treatment 115 combinations replicated 15 times. In each treatment combination, three tadpoles from 116 the same families were used to diminish intra-specific competition in accordance with 117 the principles of kin selection theory. Then, olfactory cues were provided during the 118 larval period (before Gosner stage 43) by adding 25-800 mL of water from other 119 families when changing water, minimizing maternal influence. Throughout the study 120 121 period, tadpoles were subjected to a 13L:11D photoperiod. The room temperature was maintained at  $27.72 \pm 1.12$  °C, and any temperature variations were consistent across 122 all parts of the experiment. To ensure cleanliness, we used a plastic straw to remove 123 faeces and excess food from the tanks once a day prior to feeding. In smaller 124 containers (250 ml and 500 ml), half of the water was replaced twice a week. For the 125 2000 ml containers, water replacement occurred once a week, while for the 8000 ml 126 127 containers, it was done biweekly.

When the first metamorph (Gosner stage 42) was observed, tanks were checked

daily. All metamorphs were then carefully collected and placed individually in plastic 129 vials with sand and 1 mm of water until the end of metamorphosis (Gosner stage 46). 130 We measured five variables. The length of the larval period was determined by 131 counting the number of days from the beginning of the experiment until Gosner stage 132 42. Mass at metamorphosis was measured using a balance (to the nearest 0.001 g), 133 and snout-vent length (SVL) was measured using a digital caliper (to the nearest 0.01 134 mm). Growth rate was calculated by dividing the mass at metamorphosis by the age at 135 metamorphosis (Gosner stage 42), and the survival rate was determined by calculating 136 the percentage of tadpoles that successfully underwent metamorphosis. 137

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#### 139 Data analysis

We analyzed the length of the larval period, SVL, mass at metamorphosis, and 140 growth rate using univariate two-way ANOVAs with type III sum of mean squares, 141 treating water volume, food diversity, and their interaction as fixed factors. We also 142 used a log-linear model to examine the survival rate. In cases where the univariate 143 two-way ANOVAs yielded significant results, we performed ANOVAs or one-way 144 ANOVAs with post-hoc comparisons (Fisher's LSD) to assess differences between 145 water volume or food diversity. However, given the non-significant findings from the 146 log-linear model, no further analyses were performed to investigate differences in 147 survival rates across water volume or food diversity. Statistical analyses were 148 149 conducted using IBM SPSS Statistics 20.0 (IBM Corp, Armonk, NY, USA).

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#### **RESULTS**

The effects of water volume and food diversity on the length of the larval period were significant (Table 1). However, their interaction was not significant (Table 1). Tadpoles raised at 8000 ml took less time to metamorphose than those raised at other volumes (Fisher's LSD's post hoc tests, all P < 0.001, Fig 1), while no difference was found across other water volume treatments (all P > 0.060). Tadpoles feeding on two resources reached metamorphosis earlier than those raised at single-resource treatments (P = 0.028 for all 1 of 4 LSD's post hoc tests, Fig 1).

The mass at metamorphosis was significantly influenced by water volume (Table 1, Fig. 1). However, there was no significant impact of food diversity or the interaction between water volume and food diversity (Table 1). Tadpoles reared in 8000 ml and 2000 ml had significantly larger mass at metamorphosis compared to those raised in 250 ml and 500 ml (all P < 0.001), but not between 8000 ml and 2000 ml (P = 0.807) or 250 ml and 500 ml (P = 0.992).

Water volume significantly affected SVL at metamorphosis (Table 1, Fig. 1), but food diversity, as well as water volume × food diversity interaction, did not (Table 1). The SVL of tadpoles reared at 8000 ml and 2000 ml was significantly larger at metamorphosis compared to those reared at 250 ml and 500 ml (all P < 0.003). However, there was no significant difference in SVL between tadpoles reared at 8000 ml and 2000 ml (P = 0.904), or between those reared at 250 ml and 500 ml (P = 0.777).

172 In addition to water volume (Table 1, Fig.1), food diversity, as well as the

interaction between water volume and food diversity did not significantly affect growth rate (Table 1). Tadpoles raised at 8000 ml had a greater growth rate than those raised at other treatments (all P < 0.038, Fig. 1), while tadpoles raised at 250 ml and 500 ml had a smaller growth rate than those raised at 2000 ml (both P < 0.001), but not between 250 ml and 500 ml (P = 0.809).

Water volume and food diversity did not affect survival at metamorphosis (water volume, Z = -0.97, P = 0.330; food diversity, Z = -0.57, P = 0.571), neither the interaction between water volume and food diversity (Z = 0.83, P = 0.405, Fig. 1).

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### DISCUSSION

Increasing the absolute volume of water increases the growth and developmental 183 rate of tadpoles (Gromko et al., 1973; Golay and Durrer, 1994; Smith, 1998; Durnin 184 and Smith, 2001; Kehr et al., 2014). Our study found that single tadpoles reared in 185 larger water volume had a larger body size at metamorphosis compared to those raised 186 in smaller water volume. Additionally, our results demonstrated that single B. g. 187 gargarizans tadpoles raised in larger water volume grew at a higher rate and had a 188 shorter larval period than those raised in smaller water volume. Previous studies have 189 also shown that smaller water volume decreased tadpole growth (Smith, 1998; Durnin 190 and Smith, 2001), which is consistent with our results. 191

In this study, all tadpoles were individually placed in opaque glass beakers and received the same amount of food, suggesting that the effects of water volume can occur independently of food limitation, chemical accumulation, and social

interactions. These findings support the idea that mechanisms operating in smaller 195 water volumes can lead to crowding effects. One of the mechanisms responsible for 196 the effects of water volume is an increase in the number of collisions among 197 individuals confined to limited volumes, which in turn leads to a decrease in growth 198 or development rates (John and Fenster, 1975; John and Fusaro, 1981; Rot-Nikcevic 199 et al., 2005; Fixari III et al., 2017). Nevertheless, in our experiment there was an 200 increased possibility of physical contact between tadpoles and vessel walls due to the 201 smaller water volume. Additionally, we observed higher activity levels in tadpoles in 202 smaller water volumes. This increased activity can be attributed to the increased 203 collisions and agitation, resulting in an expenditure of energy (Rugh, 1934). Crowding 204 diminishes the appetite of tadpoles and leads to slower growth (Adolph, 1931). 205 Therefore, we suggest that the frequency of physical encounters between tadpoles and 206 the vessel walls may cause psychological stress due to crowding (John and Fenster, 207 1975). 208

209 The surface area played a crucial role in restricting growth by regulating the availability of oxygen. Likewise, Yung (1885) discovered that tadpoles attained a 210 larger size in tanks with a greater surface area. In this study, the surface and bottom 211 areas of the container increase proportionally with the increase in container volume, 212 but we speculated that the oxygen levels in smaller containers may not have been 213 sufficient to restrict growth or had minimal inhibitory effects due to the increased 214 frequency of water changes as the tank volume decreased. Additionally, in tanks with 215 smaller water volumes, but with the same number of tadpoles, there was a higher 216

accumulation of fecal material on the tank bottom, which could potentially impact
growth (Podhradsky, 1932; Adolph, 1931). In this case, we employed plastic straws to
clean up feces once a day throughout the experiments, thus minimizing the potential
impact of feces on growth.

While the interaction between water volume and food diversity did not impact 221 larval growth and development, we found that food diversity plays a significant role 222 in determining the length of the larval period. Specifically, larger bodies of water with 223 two food resources resulted in a shorter larval period and earlier metamorphosis. 224 Within natural populations, tadpoles that undergo early metamorphosis can escape 225 pond drying and predators (Loman, 2002). Additionally, they acquire a larger body 226 size before hibernation, increasing their chances of survival during the longer winter 227 period (Valenzuelasanchez et al., 2015). 228

In conclusion, both water volume and food diversity had an impact on larval 229 growth and development, but these factors acted independently. The small volume of 230 water had a more pronounced effect on limiting larval growth and development 231 compared to food diversity. This could be attributed to the increased physical 232 encounters between tadpoles and vessel walls, resulting in stress from crowding or 233 psychological factors. Additionally, the crowded conditions in a small volume of 234 water could lead to limited oxygen supply, which may be a secondary factor in 235 reducing larval growth. Further studies are needed to explore the amount of available 236 237 oxygen and gain a better understanding of its role in this process.

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Response variable	Source of variation	df	MS	F-value	P-value
Length of larval period	Water volume	3	100.014	16.594	<0.001
	Food diversity	1	30.539	5.067	0.027
	Water volume × Food diversity	3	12.225	2.028	0.115
	Error	94	6.027		
SVL	Water volume	3	4.92	7.306	<0.001
	Food diversity	1	0.5	0.742	0.391
	Water volume × Food diversity	3	0.532	0.79	0.502
	Error	94	0.673		
Body mass	Water volume	3	0.015	19.447	< 0.001
C C	Food diversity	1	< 0.001	0.044	0.835
8	Water volume × Food diversity	3	0.0004	0.561	0.642
	Error	94	0.001		
Growth rate	Water volume	3	20.244	32.709	< 0.001
	Food diversity	1	0.391	0.631	0.429
	Water volume × Food	3	0.46	0.743	0.529

Table 1. Two-way ANOVA tables for the effects of water volume and food diversityon metamorphic traits in a *Bufo gargarizans* population.

	diversity					
	Error	94	0.619			
354						
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Fig. 1. Influences of water volume and food diversity on (A) age at metamorphosis,
(B) SVL, (C) body mass, (D) growth rate and (E) survival of *Bufo gargarizans gargarizans* (black circles, two resources; open circles, single resource; Error bars
indicate standard errors).

