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**Study of the anatomy and histology of the female reproductive system of the Asian
snake-eyed skink, *Ablepharus pannonicus* (Sauria: Scincidae)**

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Abstract. This study examines and contrasts the anatomy and histology of the female reproductive system in *Ablepharus pannonicus*, a member of the Scincidae family, across the spring and autumn seasons. Female specimens of *A. pannonicus* were collected from the northern slopes of the Shorkhah Dizah village (34°23'49.1"N 46°03'15.1"E), located 125 km west of Kermanshah Province in western Iran. After anaesthesia, the specimens were dissected in the laboratory, and their reproductive systems were processed for tissue analysis, using serial sections stained with Hematoxylin and Eosin for detailed histological examination. The findings indicate that during the spring, the ovaries of *A. pannonicus* contain all stages of follicular development, including primordial, mature, and ovulation-ready follicles. In contrast, the autumn samples predominantly exhibit primordial or preovulatory follicles, with a notable absence of mature follicles. These observations suggest a seasonal reproductive pattern in female *A. pannonicus*, characterized by significant morphological changes and heightened activity within the ovaries and genital tracts during spring. As the ambient temperature drops and the reproductive season concludes, gonadal size and activity decrease, leading to the absence of reproductive activity in autumn. It is hypothesized that during this period, the animals engage in feeding and fat storage in preparation for hibernation.

Keywords. reproductive cycle, *Ablepharus pannonicus*, reproductive system, ovary

The exploration of reproductive diversity among lizards offers a rich tapestry of biological phenomena for scientific inquiry. These reptiles display a remarkable array of reproductive strategies: from oviparity to viviparity, prolific to sparse offspring production, and varied levels of parental investment. Some species are known to lay eggs immediately post-shell formation, while others retain shelled eggs for prolonged periods before laying. Remarkably, viviparity has independently evolved numerous times within squamates, and certain species even exhibit parthenogenesis. These reproductive variations offer a glimpse into the captivating complexity of lizard reproductive ecology (Rheubert et al., 2014).

Lizards typically exhibit one of three reproductive cycles: constant, associated, or dissociated. The constant cycle is characterized by nearly year-long gonadal activity. In contrast, the associated and dissociated cycles feature a discontinuous mating season. The associated cycle is marked by a surge in gonadal activity just before the mating season, synchronously in both sexes, obviating the need for sperm storage due to its ready availability (Censky, 1995; Huang, 1997). Conversely, the dissociated cycle is defined by diminished gonadal activity during the mating season, with a peak in the non-mating period. Notably, male gonadal activity is briefer compared to females, necessitating the storage of sperm within the female reproductive tract for subsequent fertilization (Torki, 2006).

The oviductal structure in lizards exhibits a conserved morphology, characterized by three distinct regions identifiable across species. These regions, extending from the caudal to the cranial end, comprise the nonglandular uterus—commonly referred to as the vagina—the glandular uterus, typically known as the uterus, and the infundibulum (Siegel et al., 2011). Female lizards possess bilateral oval ovaries anchored to the dorsal body wall via a slender mesovarium. The ovarian cortex is surrounded by the tunica albuginea, and is lined by a simple squamous epithelium (Klosterman, 1983; Guraya, 1989). Oogenesis occurs within the germinal beds situated dorsally on the ovary, proximal to the mesovarium. These beds harbor

proliferating oogonia, undifferentiated somatic cells, oocytes encased by a few somatic cells, and primary follicles (Jones and Guillette, 1982; Klosterman, 1983). The ovarian follicles, serving as the functional units of the ovary, are organized into a hierarchical structure, including primordial, primary, secondary, previtellogenic, vitellogenic, and preovulatory follicles. This follicular hierarchy is a well-defined feature of reptilian ovarian architecture (Guraya, 1989; Etches and Petitte, 1990).

The skink *Ablepharus pannonicus* (see Fig. S1) inhabits a range of environments including semi-desert and desert areas, steppe habitats, and mountainous terrains. Its geographical distribution extends from the Middle East to Central and South Asia, covering countries such as Iraq, Iran, Jordan, Syria, Afghanistan, Pakistan, India, Kyrgyzstan, and Uzbekistan (Karamiani, 2018). Despite the breadth of studies on various aspects of this species, there is a notable gap in the literature regarding the anatomical and histological characterization of the female reproductive system. Addressing this lack of information, the present study provides the first detailed description of the anatomy and histology of the female reproductive system of *A. pannonicus*, offering insights into a previously unexplored aspect of this widely distributed lizard.

This research was conducted in the natural surroundings of the village of Shorkhah Dizah (34°23'49.1"N 46°03'15.1"E), located 125 km west of the city of Kermanshah, in Kermanshah Province, western Iran. Sampling was conducted in the spring (April–May) and autumn (October–November) of 2021, with four adult female specimens of *A. pannonicus* collected in each season. These two sampling periods were selected to capture contrasting reproductive stages: spring representing peak ovarian activity and autumn reflecting a regressed state, consistent with seasonal reproductive patterns reported in scincid lizards (Sever and Hopkins, 2004; Vergilov et al., 2018). According to long-term climate normals (1981–2010) reported by the Iran Meteorological Organization (IRIMO), average temperatures in the study area range

from 15–22 °C in spring and 9–17 °C in autumn. Rainfall is moderate in spring but substantially lower in autumn (Iran Meteorological Organization, 2021).

Upon arrival at the laboratory, the specimens underwent anaesthesia using diethyl ether-soaked cotton. Subsequently, we euthanised them through intracoelomic injection of sodium pentobarbital, following the animal care guidelines approved by Southeastern Louisiana University Animal Care and Use Committee (Rheubert et al., 2020). Next, we meticulously recorded morphometric parameters, including snout–vent length (SVL) and body mass. A longitudinal incision was made on the ventral surface of the specimens, allowing for the removal of the digestive tract. This surgical approach exposed the reproductive system, which we then photographed for subsequent analysis. The entire oviductal tract was carefully dissected and weighed using a digital analytical balance. Using this data, we calculated the gonadosomatic index (GSI), providing a quantitative measure of reproductive investment (Jacobson, 2007). The GSI was calculated as $GSI = (\text{ovary weight} / \text{body weight}) \times 100$, reflecting the proportional reproductive investment of each individual. After the oviductal tract was removed from the lizard's body, the samples were immediately placed in 10% formalin for 72 h. Subsequently, the samples were dehydrated in a series of ethanol solutions: 60% for 60 min, 70% for 30 min, 80% for 30 min, and 96% for 120 min. For clearing, the samples were placed in three containers containing xylene, each for 30 min. The samples were then embedded in paraffin in an incubator set at 58°C. In this step, three containers of paraffin were used, and the capsules containing the tissue samples were placed in each container for 120 min to allow the paraffin to replace the xylene in the tissue. L-shaped metal molds, known as Leuckhard molds, were used for molding. The mold was adjusted to the appropriate size, and molten paraffin was then poured into the mold. The samples were subsequently placed vertically inside it. In the next step, the blocks were separated from the mold, resulting in a molded block for each individual. A rotary microtome (CUT SLEE 4060) was used to obtain transversal and

longitudinal sections from paraffin blocks containing the entire oviductal tract. Sections were prepared from multiple regions along the tract, and representative slides were selected for histological analysis. The sections were stained using hematoxylin-eosin staining (Suvarna et al., 2018).

The female reproductive system in *Ablepharus pannonicus* comprises a pair of ovaries and an oviduct. The oviducts traverse past the kidneys and terminate in the cloaca (Fig. 1A, B). The ovaries are situated within the abdominal cavity as whitish glandular structures, attached to the dorsal wall by the mesovarium. The ovarian surface exhibits small and large protrusions due to the presence of follicles at various stages of maturity. Each ovary is enveloped by a thin covering known as the tunica albuginea. During the spring, all types of follicles (especially vitellogenic and preovulatory follicles) are observable in the ovarian sections (Fig. 1C). However, in autumn samples, mature follicles are scarce, and the predominant follicles are in the primary, secondary, or previtellogenic stage (Fig. 1D).

Microscopic examination of the previtellogenic follicle wall reveals at least two distinct cell types—large and small cells (see Fig. S2). The ovarian and body weight data, and gonadosomatic index data collected from each season are presented in Table S1. No notable structural differences were observed in the oviductal anatomy or histology between spring and autumn specimens. Moving along the oviduct, the most anterior portion is referred to as the infundibulum. The epithelial cells lining its proximal region exhibit a squamous to cuboidal morphology. Toward the distal region, the cellular height increases, culminating in a cylindrical form. Notably, certain cells in the distal infundibulum display abundant cilia (Fig. 2A, B).

The glandular uterus constitutes the central segment of the oviduct in *A. pannonicus*. Its wall is densely populated with mucous glands, which are acinar in configuration and embedded within the tissue. The predominantly columnar cells of these glands play a crucial role in eggshell formation through their secretions (Fig. 2C, D).

The most posterior region of the oviduct is the non-glandular uterus, connecting the glandular uterus to the urodeum of the cloaca. In this area, numerous crypts are present, their abundance increasing as one approaches the cloacal urodeum (see Fig. 3 and Fig.S3).

Unlike the glandular uterus, this region lacks secretory glands, allowing for clear demarcation (see Fig. 4). The number of Germinal Beds (GBs) per ovary exhibits both intra- and inter-specific variation, ranging from one to six among lizard species (Jones et al., 1979; Radder et al., 2008). Studies have posited a correlation between the number of GBs per ovary, clutch size, and breeding frequency. Jones and Guillelte (1982) identified a pattern linking the number of GBs to clutch size. Typically, species that undergo monoallochronic ovulation—ovulating a single egg alternately from each ovary, as seen in the Dactyloidae family (Jones et al., 1979)—and those with low fixed clutch sizes and monoautochronic ovulation—releasing a single egg simultaneously from both ovaries, such as in Gekkota (Jones and Summers, 1984) and Gymnophthalmidae (Vitt, 1982)—possess one GB per ovary. Conversely, species with high fecundity, characterized by polyautochronic ovulation and multiple large clutches, exhibit two or more spatially distinct GBs in each ovary (Radder et al., 2008). In all *A. pannonicus* specimens examined during the spring, GBs were indistinct. However, the presence of a single follicle poised for ovulation in both ovaries suggests a monoallochronic ovulation pattern.

Within the walls of previtellogenic follicles in certain lizards, granulosa cells of varying types are discernible. For instance, in *Ablepharus kitaibelii*'s previtellogenic follicles, three granulosa cell types—small, large, and pyriform—are identifiable (Vergilov et al., 2018). In this study, two cell types—large and small—were observed in *A. pannonicus*'s previtellogenic follicle walls, yet pyriform cells were absent.

In species like *Eumeces egregius* (Schaefer and Roeding, 1973) and *Scincella lateralis* (Sever and Hopkins, 2004), the non-glandular uterus contains numerous ciliated crypts functioning as sperm storage sites. Examination of the non-glandular uterus of *A. pannonicus*

revealed numerous crypts, similar to those reported in the species mentioned above. However, no sperm was detected in specimens collected during either spring or autumn. This absence may reflect phylogenetic differences among scincid lizards or limitations of our sample size, rather than conclusively indicating that the crypts are not used for sperm storage.

Glandular uterine glands play a pivotal role in egg membrane formation via their secretions. These glands exhibit considerable morphological diversity across lizard species. In *Plestiodon obsoletus*, the glands are tubular (Guillette Jr. et al., 1989), while in *Chalcides chalcides*, they are tubular with simple cuboidal epithelium (Blackburn et al., 1998), and in *Chalcides ocellatus*, they are simple alveolar (Corso et al., 2000). In these species, secretions are expelled through an end duct to the uterus's glandular lining. The glands in the glandular uterine wall of *A. pannonicus* were acinar with columnar epithelial cells in specimens from both spring and autumn. While no seasonal structural differences were noted, potential variations in functional activity cannot be ruled out.

The infundibulum, featuring numerous crypts in lizards like *Acanthodactylus scutellatus* and *Hemidactylus turcicus*, is recognized as a sperm storage site (Bou-Resli et al., 1981; Eckstut et al., 2009). However, comprehensive documentation of sperm storage in this region among skinks is lacking. In *A. pannonicus*, as in other skinks, sperm was not detected in the infundibulum.

These findings suggest that reproduction in female *A. pannonicus* is seasonal, as indicated by significant morphological changes in the gonads and increased activity in the genital ducts during spring. This conclusion is further supported by a marked increase in the gonadosomatic index (GSI) observed in spring-collected specimens compared to those collected in autumn, reflecting a higher reproductive investment during the active breeding season. As the ambient temperature declines, a noticeable seasonal reduction in both gonadal size and GSI is evident, corresponding with the cessation of reproductive activity. Consequently, this lizard exhibits no

signs of sexual activity during the autumn season. This seasonal variation in gonadal condition and reproductive readiness aligns with regional climatic patterns, where average daily temperatures range from 15–22 °C in spring and decrease to 9–17 °C in autumn in the mountainous areas south of Kermanshah Province.

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SUPPLEMENTARY MATERIAL

Supplementary material associated with this article can be found at <http://www.unipv.it/webshi/appendix> > Manuscript number 16334

- 218 Blackburn, D. G., Kleis-San Francisco, S., Callard, I. P. (1998): Histology of abortive egg sites
219 in the uterus of a viviparous, placentotrophic lizard, the skink *Chalcides chalcides*. J Morphol.
220 **235**: 97-108.
- 221 Bou-Resli, M. N., Bishaw, L. F., Al-Zaid, N. S. (1981): Observations on the fine structure of
222 the sperm storage crypts in the lizard *Acanthodactylus scutellatus hardyi*. Arch. Biol. (Brux.)
223 **92**: 287–298.
- 224 Censky, E. J. (1995): Reproduction in two Lesser Antillean populations of *Ameiva plei*
225 (Teiidae). J. Herpetol. **29**: 553-560.
- 226 Corso, G., Delitala, G. M., Carcupino, M. (2000). Uterine morphology during the annual cycle
227 in *Chalcides ocellatus tiligugu* (Gmelin) (Squamata: Scincidae). J. Morphol. **243**: 153-165.
- 228 Eckstut, M. E., Lemons, E. R., Sever, D. M. (2009): Annual dynamics of sperm production and
229 storage in the Mediterranean Gecko, *Hemidactylus turcicus*, in the southeastern United States.
230 Amphib-Reptil. **30**: 45-56.
- 231 Etches, R. J., Petite, J. N. (1990): Reptilian and avian follicular hierarchies: models for the
232 study of ovarian development. J. Exp. Zool. **256**: 112-122.
- 233 Guillette Jr, L. J., Fox, S. L., Palmer, B. D. (1989): Oviductal morphology and egg shelling in
234 the oviparous lizards *Crotaphytus collaris* and *Eumeces obsoletus*. J. Morphol. **201**: 145-159.
- 235 Guraya, S. S. (1989): Ovarian follicles in reptiles and birds. Zoophysiology **24**: XIII-285.
- 236 Huang, W. S. (1997): Reproductive cycle of the oviparous lizard *Japalura brevipes*
237 (Agamidae: Reptilia) in Taiwan, Republic of China. J. Herpetol. **31**: 22-29.

238 Iran Meteorological Organization. 2021. Climate Normals (1981-2010). Iran Meteorological
 239 Organization, Tehran, Iran.

240 Jacobson, E.R. (2007): Biology, Medicine, and Surgery of Reptiles. Boca Raton, Florida, CRC
 241 Press.

242 Jones, R.E., Fitzgerald, K.T., Duvall, D., Banker, D. (1979): On the mechanisms of alternating
 243 and simultaneous ovulation in lizards. *Herpetologica* **35**: 132-139.

244 Jones, R. E., Guillette Jr, L. J. (1982): Hormonal control of oviposition and parturition in lizards.
 245 *Herpetologica* **38**: 80-93.

246 Jones, R. E., Summers, C. H. (1984): Compensatory follicular hypertrophy during the ovarian
 247 cycle of the house gecko, *Hemidactylus frenatus*. *Anat. Rec.* **209**: 59-65.

248 Karamiani, R., Rastegar-Pouyani, N., Rastegar-Pouyani, E. (2018): Modeling the past and
 249 current distribution and habitat suitability for *Ablepharus grayanus* and *A. pannonicus* (Sauria:
 250 Scincidae). *Asian Herpetol. Res.* **9**: 56-64A.

251 Klosterman, L. L. (1983): The ultrastructure of germinal beds in the ovary of *Gerrhonotus*
 252 *coeruleus* (Reptilia: Anguidae). *J. Morphol.* **178**: 247-265.

253 Radder, R. S., Pizzatto, L., Shine, R. (2008): Morphological correlates of life-history variation:
 254 is lizard clutch size related to the number of germinal beds in the ovary? *Biol. J. Linn. Soc.* **94**:
 255 81-88.

256 Rheubert, J. L., Siegel, D. S., Trauth, S. E. (2014): Reproductive biology and phylogeny of
 257 lizards and tuatara. Boca Raton, Florida, CRC Press.

258 Rheubert, J., Pasternak, M. A., Ely, M., Siegel, D. S., Trauth, S. E., Gribbins, K. M., Sever, D.
 259 M. (2020): Seasonal histology and ultrastructure of the urogenital system in two sympatric
 260 lizards. *J. Zool.* **310**: 273-286.

261 Schaefer, G. C., Roeding, C. E. (1973): Evidence for vaginal sperm storage in the mole skink,
 262 *Eumeces egregius*. *Copeia* **1973**: 346-347.

263 Sever, D. M., Hopkins, W. A. (2004): Oviductal sperm storage in the ground skink *Scincella*
 264 *laterale* Holbrook (Reptilia: Scincidae). *J. Exp. Zool. A Comp. Exp. Biol.* **301**: 599-611.

265 Siegel, D. S., Miralles, A., Chabarria, R. E., Aldridge, R. D. (2011): Female reproductive
 266 anatomy: cloaca, oviduct, and sperm storage. In: Reproductive biology and phylogeny of
 267 snakes, pp. 347-409. Aldridge, R. D., Sever, D. M., Eds, Florida, CNC Press

268 Suvarna, K. S., Layton, C., Bancroft, J. D. (2018): Bancroft's theory and practice of histological
 269 techniques. Elsevier Health Sciences.

270 Torki, F. (2006): Spermatogenesis in the agama *Trapelus lessonae* (Agamidae: Reptilia) in the
 271 central Zagros Mountains, Iran. *Zool. Middle East* **38**: 21-28.

272 Vergilov, V. S., Necheva, V. G., Zlatkov, B. P. (2018): Reproduction of Snake-eyed Skink
 273 *Ablepharus kitaibelii* (Bibron & Bory de Saint-Vincent, 1833) (Squamata: Scincidae) in
 274 Bulgaria. *Acta Zool. Bulg.* **70**: 507-516.

275 Vitt, L. J. (1982): Sexual dimorphism and reproduction in the microteiid lizard,
 276 *Gymnophthalmus multiscutatus*. *J. Herpetol.* **16**: 325-329.

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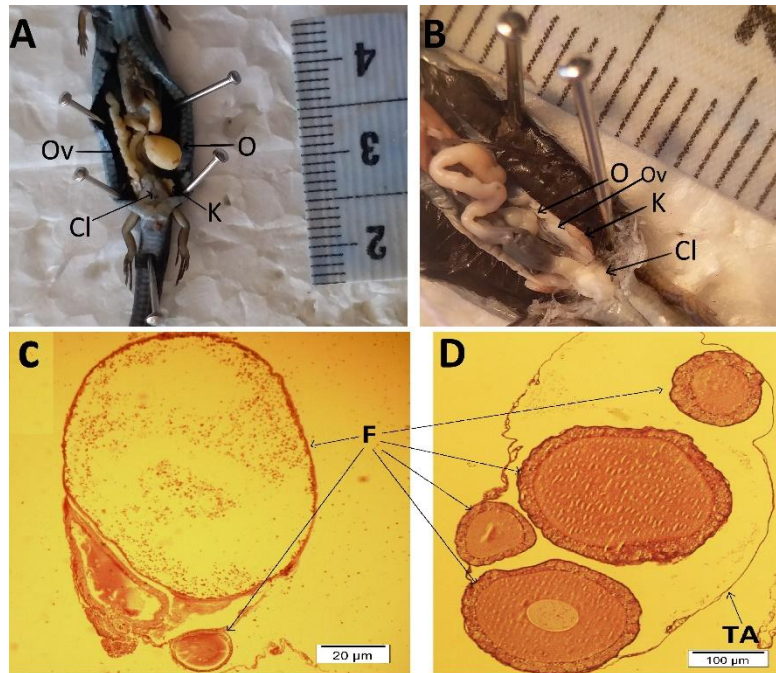
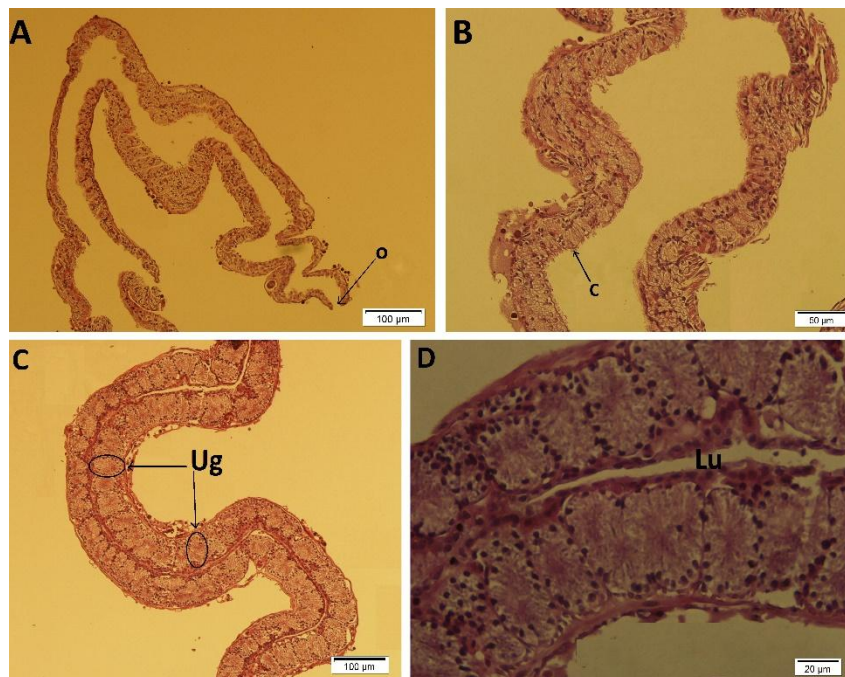


Fig. 1. Gross anatomy and ovarian histology of the female reproductive system of *Ablepharus pannonicus* in two seasons. (A, B) Dissected views of the female reproductive system in spring (A) and autumn (B), showing the position of the ovaries and oviducts relative to the kidneys and cloaca. (C) Section from a spring-collected individual showing an advanced follicle (likely preovulatory) and developing follicles (presumed vitellogenic). (D) Section from an autumn-collected individual illustrating follicles at various earlier stages of development, including presumed early, intermediate, and previtellogenic follicles. Sections stained with hematoxylin and eosin (H&E). Abbreviations: O, ovary; Ov, oviduct; K, kidney; Cl, cloaca; F, follicle; TA, tunica albuginea.



296

297 **Fig. 2.** Longitudinal histological sections of the infundibulum and glandular uterus in
298 *Ablepharus pannonicus*. (A) Low magnification view showing the entire infundibulum from
299 the anterior ostium to the posterior connection with the uterus. (B) Higher magnification of the
300 posterior region of the infundibulum, revealing epithelial cilia. (C) Low magnification
301 longitudinal section of the glandular uterus, showing the distribution of uterine glands within
302 the uterine wall. (D) Higher magnification view of the glandular uterus, illustrating the acinar
303 morphology of the uterine glands. Sections stained with hematoxylin and eosin (H&E).
304 Abbreviations: O, ostium; C, cilia; Ug, uterine gland; Lu, lumen.

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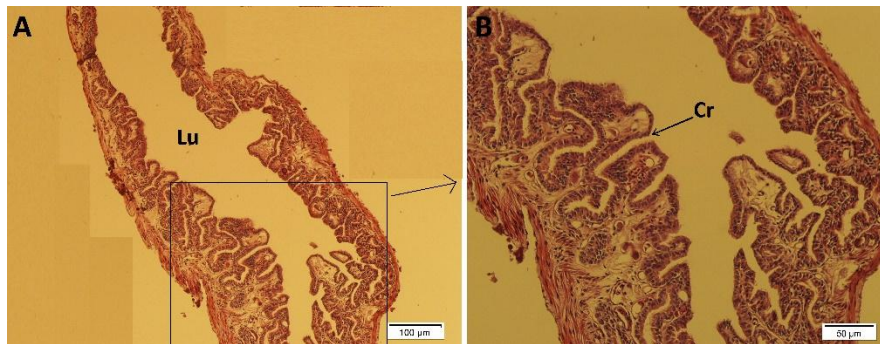
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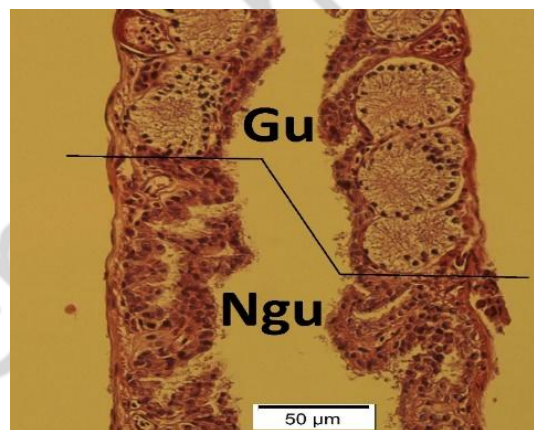
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311 **Fig. 3.** Longitudinal section of the non-glandular uterus (vagina) of *A. pannonicus* collected in
312 spring. (A) Low magnification showing overall tissue structure and distribution of epithelial
313 crypts. (B) Higher magnification illustrating the detailed morphology of crypts. Sections stained
314 with hematoxylin and eosin (H&E). Abbreviations: Lu: lumen, Cr: crypt.

315



316

317 **Fig. 4.** Longitudinal section illustrating the junction between the glandular (Gu) and non-
318 glandular (Ngu) regions of the uterus in *A. pannonicus*. The boundary between the two is
319 marked by a black line. Section stained with hematoxylin and eosin (H&E). Abbreviations: Gu:
320 glandular uterus, Ngu: non-glandular uterus.

321