

Potassium silicate enhances drought tolerance of *Bellis perennis* by improving antioxidant activity and osmotic regulators

A. Oraee, A. Tehranifar^(*), Z. Ghorbani, P. Sayad-Amin

Department of Horticultural Science and Landscape, Ferdowsi University of Mashhad, Iran.

Key words: Antioxidant activity, fertilizer, osmotic regulation, secondary metabolites.

Abstract: Ornamental plants can usually encounter various types of environmental stress, which reduce plant productivity. A proper application of fertilizers can improve plants' tolerance to drought stress. Nutrients such as potassium and silicon are known to have beneficial effects. This study aimed to evaluate the growth of Bellis perennis under drought stress (80, 70, and 60% FC) and with the application of potassium silicate (0, 2, and 4 mM). The results showed that potassium silicate (2 and 4 mM) increased K and Si accumulation in plants under drought stress. Plants treated with potassium silicate under drought stress exhibited a lower degree of electrolyte leakage and less MDA accumulation in the following order: 2 and 4 mM potassium silicate. An increase in relative water content and chlorophyll was observed with application of potassium silicate under drought stress. Regardless of potassium silicate, the plant enzymatic defense system was significantly improved compared to non-stressed plants. Potassium silicate enhanced the amount of osmotic regulators (carbohydrate and proline) and secondary metabolites (flavonoids and phenols) compared to control plants regardless of drought stress. The anthocyanin content in the flowers significantly decreased by 32.2% when the plants were treated with 4 mM potassium silicate at 60% FC, compared to 80% FC. In conclusion, potassium silicate mitigated the effects of drought stress, enhanced plant tolerance to drought stress, increased the activity of antioxidant enzymes, and improved the amounts of osmotic regulators and secondary metabolites.

1. Introduction

Plant growth is usually affected by numerous abiotic stressors (Calanca, 2017). Deficits in water supplies are a major environmental threat, currently affecting more than 41% of the world's land mass. Projections for 2050 show a further increase in the magnitude and impact of this environmental threat (Prăvălie, 2016). As global temperatures increase, the annual maximum temperature is estimated to increase by 5°C per year by the end of the 22nd century. This problem will lead to



(*) **Corresponding author:** tehranifar@um.ac.ir

Citation:

ORAEE A., TEHRANIFAR A., GHORBANI Z., SAYAD-AMIN P., 2023 - Potassium silicate enhances drought tolerance of Bellis perennis by improving antioxidant activity and osmotic regulators. - Adv. Hort. Sci., 37(4): 377-389.

Copyright:

© 2023 Oraee A., Tehranifar A., Ghorbani Z., Sayad-Amin P. This is an open access, peer reviewed article published by Firenze University Press (http://www.fupress.net/index.php/ahs/) and distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement:

All relevant data are within the paper and its Supporting Information files.

Competing Interests: The authors declare no competing interests.

Received for publication 18 May 2023 Accepted for publication 21 August 2023 more frequent and extreme droughts in many parts of the world (Gao *et al.*, 2020). Such environmental types of stress significantly cause a decline in crop yields falling below maximum potential (Raza *et al.*, 2020). Environmental stress can suppress plant growth by interrupting various processes such as cell metabolism, nutrient uptake, and maintenance of turgor pressure (Kusvuran and Dasgan, 2017).

Regarding the proper use of fertilizers for improving plant tolerance to drought stress, the application of nutrients can be largely beneficial. Such applications have reportedly included potassium and silicon, with essential functions in plant metabolism (Bukhari et al., 2020; Ibrahim et al., 2020). Potassium is a vital fertilizer involved in numerous biochemical and physiological processes, including stress tolerance, plant growth, yield, and quality. Potassium (K) is necessary for many physiological processes, for example, maintaining turgor, translocation of photosynthetic substances to sinking organs, activation of enzymes, synthesis of proteins, transport of solutes in the phloem, and maintenance of cation-anion balance in the cytosol and vacuole. Furthermore, K reportedly facilitates osmoregulation, stomatal movements, and tropism, while it is mainly absorbed from the soil through the roots (Qi et al., 2019). However, drought stress causes a reduction in the absorption of elements. The reduced sensitivity of K-deficient plants to drought stress is related to several factors. These factors include the role of K in regulating stomata stomatal and water balance, as well as osmotic potential in vacuoles (Haworth et al., 2018). Applying potassium fertilizer mitigates the adverse effects of these stressors on plant growth (Qi et al., 2019).

Silicon (Si), the second most abundant element on Earth, can increase plant tolerance to biotic and abiotic stressors such as frost, heat, pests, drought, diseases, and nutrient imbalance (Wang et al., 2021). Si deficiency reportedly decreased photosynthesis, also increased disease incidence, insect infestation, wilting, and postharvest decline. while all of these symptoms are signs of stress (Reynolds et al., 2009; Dallagnol et al., 2012; Weerahewa et al., 2015). Si usually contributes to healthy plant development and is essential for cell development and differentiation. The protective role of Si in drought conditions is mainly associated with an enhanced level of water retention, which promotes photosynthesis (Zhang et al., 2018). It accelerates the accumulation of osmolyte regulators (proline and carbohydrates) as well as antioxidant activities in plants exposed to

stress in the environment (Moussa and Shama, 2019). Potassium silicate (K-silicate) is used as a source of highly soluble K and Si. K-silicate does not contain volatile organic compounds, and its application does not result in the release of hazardous or pollutant byproducts (Romero-Aranda *et al.*, 2006). In a relevant study, the application of K-silicate to the soil, for several plant species under irrigation with water-deficit conditions, resulted in the highest biomass of all species (Moussa and Shama, 2019). Si can act as a growth regulator and can potentially increase plant growth under drought stress. Spraying K-silicate and other nanomaterials can potentially reduce the adverse effects of drought stress on crops (Zahedi *et al.*, 2020).

The common daisy (*Bellis perennis*) belongs to the Asteraceae family and is known as an archetype. *Bellis perennis* is an important ornamental and medicinal plant with a global distribution. It is one of the first flowering species and is a crucial member of spring bloomers (Siatka and Kašparová, 2010). Daisies are grown for their beauty, either for color or aesthetic reasons. They are naturally able to provide economic, environmental, and social benefits. However, water-deficit largely affects the ornamental value of this species.

In the current study, we focused on the response of daisies to drought stress, while monitoring their antioxidant enzymes, substances for osmotic regulation, and secondary metabolites. Although many studies have already considered the role of potassium silicate in reducing the adverse effects of drought stress in various plants, there is little information about the effects of potassium silicate on ornamental plants under drought-stress conditions. Therefore, the objective of the current study was to investigate the effects of potassium silicate on the characteristics of daisies under drought-stress conditions. The mechanisms of action by potassium silicate, their advantages for ornamental plants, and their ability to create drought tolerance provide a scientific basis for using potassium silicate to alleviate drought stress.

2. Materials and Methods

Plant culture, drought stress, and potassium silicate treatments

This study was carried out at the Ferdowsi University of Mashhad (autumn-spring 2021). Seeds of *Bellis perenesis* L. were purchased from Takii seed company. In September 2021, the seeds were grown in polyethylene bags containing a mixture of peat and perlite (3:1) for four weeks (trifoliate stage) under controlled conditions (21°C/17°C day/night and 45-55% humidity under 100 mmol photons m-2 s-1. Four weeks later, the trifoliate seedlings were transplanted into pots. For each treatment, the experiment was laid out with three pots. The pots were placed in a greenhouse (air temperature of 21±2°C and relative humidity of 62±2%) during the growing periods. Irrigation started after one day, and the plants were well watered for a few months (90-85% FC). Drought treatment was initiated by omitting irrigation, and potassium silicate (K,SiO₃) was used as the Si source. Potassium was administered to the plants in the form of liquid potassium silicate (K₂SiO₂) (10% K₂O, 25% SiO₂) at three concentrations (0, 2, 4 mM) (The concentrations of potassium silicate were selected according to a pretest). In March, the plants were treated with different solutions, i.e. (1) 1/2 Hoagland's solution without the addition of K₂SiO₂, (2) 1/2 Hoagland's solution with the addition of 2 mM K₂SiO₃, (3) 1/2 Hoagland's solution with the addition of 4 mM K₂SiO₃. Potassium silicate was applied as a treatment for one month in March, and irrigation treatments began with drought stress (80%, 70%, and 60% FC) in April. Three levels of water deficit (i.e. 80, 70, and 60% of field capacity, FC) were applied from April to June. The gravimetric method (Campbell and Mulla, 1990) was used for irrigation for two months. First, several pots were completely irrigated so that the water permeated all pores in the soil. Then, the pots were wrapped with plastic covers to prevent evaporation and transpiration. The pots were weighed until their weight remained constant for two consecutive measurements. Then, a soil sample was taken to the laboratory. The fresh weight was measured and the dry weight was calculated after 12 hours of storage in an oven at 105°C. The percentage of moisture content by weight, required for suitable crop production, is calculated based on the following equation:

FC = (A-B / B) × 100

where FC, A, and B are the field capacity, the weight of moist soil after gravity drainage, and the weight of the sample dried at 105°C for 12 hours, respectively.

The weight difference between water-saturated and oven-dried soil was taken as the weight of water needed to bring the pots to field capacity, and then lower water contents in the soil (% field capacity) were calculated accordingly. During the period of treatments, the pots were regularly weighed, and additional water was supplied when necessary. For each test, there were three replicates containing five plants, making a total of 15 plants. At the end of the experiment, the fresh leaves were used for measuring electrolyte leakage, RWC, and chlorophyll content. For determining the nutrition concentration and proline content, dried leaves were frozen in liquid nitrogen and stored at -80°C until the time of measurements.

Determination of K and Si concentration

The K and Si concentrations were determined on the dry leaves samples. Oven-dried leaves (300 mg of the dried samples) were weighed and burned in a muffle furnace at 550°C for 8 hours. The K concentration was determined by flame photometry (PFP7, Jenway, UK). The Si concentration was determined by the colorimetric ammonium vanadate method (Jaiswal, 2003).

Measurement of electrolyte leakage and relative water content

Electrolyte loss was determined according to a method used by Gusta *et al.* (2003), and relative water content (RWC) was calculated via a method used by Pieczynski *et al.* (2013).

Measurement of antioxidant activity and malondialdehyde (MDA)

Antioxidant activity was determined using 1,1diphenyl-2-picrylhydrazyl (DPPH). The extract (100 mg of fresh weight + ethanol) was blended with 960 µL of DPPH in methanol. The supernatant was centrifuged for 5 min and kept in the dark room. The DPPH was determined using a Shimadzu UV-1800 spectrophotometer at 515 nm (Kedare and Singh, 2011). Malondialdehyde (MDA) content was measured according to Velikova et al. (2000) methods. Leaf tissues (0.5 g of each) were homogenized in 8 ml of 0.1% (w/v) trichloroacetic acid and the homogenates were centrifuged for 10 min at 4 °C, after which the supernatants were used for malondialdehyde analysis. Equal volumes of extracts were mixed with 0.5% (w/v) of thiobarbituric acid made in 5% (w/v) trichloroacetic acid and heated at 100°C water bath for 20 min, after which their actions were stopped in the ice bath. After centrifuging, the absorbance of the supernatant was measured at 450, 532, and 600 nm.

Determination of antioxidant enzyme activities

Extraction was performed according to a method used by DaCosta and Huang (2007). Samples (0.5 g of fresh weight) were ground in liquid nitrogen and were homogenized in 4 mM phosphate buffer (pH 7.8), 60 M riboflavin, 195 mM methionine, 3 M EDTA, and 1.125 mM nitro blue tetrazolium chloride (NBT). Enzyme activities were expressed per fresh weight of the sample. One SOD activity unit was defined as the amount of enzyme required to cause 50% inhibition of nitro blue tetrazolium chloride (NBT) photoreduction (Sairam et al., 2002). Catalase activity (CAT) was measured as described by Abedi and Pakniyat (2010). The reaction solution consisted of 50 mM K-phosphate buffer (pH 7.0), ten mM H₂O₂, and 50 mL enzyme extract. The decomposition of H₂O₂ was measured at 240 nm. The peroxidase activity (POD) was measured by the guaiacol method (Guan et al., 2015). The oxidation of guaiacol was monitored by observing changes in the absorbance values at 470 nm for 3 min. The reaction mixture contained 50 ml of 100 mM PBS (pH 6.0), 10 mM H₂O₂, 2.58 mM of guaiacol. The reaction was started by adding the enzyme extract to the reaction mixture solution.

Measurement of photosynthetic pigments

Chlorophyll contents (Chl a, b, and total Chl) were measured by squashing the leaves (200 mg) in 10 ml 80% acetone solution, and the chlorophyll content was determined at 645 and 663 nm, respectively using a Shimadzu UV-1800 spectrophotometer (Nagata and Yamashita, 1992).

Determination of osmotic regulators

Carbohydrates were determined using the Anthrone reagent method. Fresh leaves (500 mg) were placed in 70% methanol and reached the required volume with distilled water. The samples were used for estimations of carbohydrate content using the Anthrone reagent (McCready *et al.*, 1950). Proline content was calculated according to Bates *et al.* (1973). The leaf extract (0.1 mg leaf sample + 10 ml sulfosalicylic acid) was homogenized in glacial acetic acid and ninhydrin acid. Then, the solution was heated in a boiling water bath. After cooling, 5 mL of toluene was added, and then the top layer of the solution was removed and centrifuged at 3000 g for 5 minutes. The proline content was determined at 520 nm using a Shimadzu UV-1800 spectrophotometer.

Secondary metabolite measurements

Total phenolic content was measured using the Folin-Ciocalteu reagent method (Singleton and Rossi,

1965). In the Folin-Ciocalteu method, 250 µl of the alcoholic extract (100 mg + 10 ml ethanol) was diluted to a known volume with distilled water, 10% Folin reagent, and 7.5% sodium carbonate. The phenolic content was determined at 675 nm using a Shimadzu UV-1800 spectrophotometer. Assaying the total anthocyanin content followed, a method by Sukwattanasinit et al. (2007), where two buffer solutions were used (25 mM K-chloride pH 1.0 and 0.4 M Na-acetate pH 4.5). The values were noted at 510 nm using a Shimadzu UV-1800 spectrophotometer. Flavonoid content was assayed according to a method by Zou et al. (2004). The extract (500 mg + 5 ml ethanol) was homogenized in 4.5 mL distilled water and 0.3 mL 5% NaNO₂. Next, after mixing the solution properly, 1 mL of 10% AlCl₂-6H₂O, 2 mL of 1 M NaOH, and distilled water were added to the reaction mixture. The absorbance values were determined at 510 nm using a spectrophotometer (Shimadzu UV-160A).

Statistical analysis

The difference between treatments was determined using a factorial layout and a completely randomized experimental design with three replicates followed by the LSD testing (P<0.01). Data were subjected to two-way analysis (ANOVA) with repeated measures and were analyzed using the SAS statistical package (version 9.2, SAS Institute, Cary, NC, USA).

3. Results

The potassium and silicon concentrations were significantly (*P*<0.01) affected by fertilizer and drought stress. Potassium (K) concentration in the control plants decreased by 14.9% under drought stress at 60% FC compared to 80% FC. A decrease in leaf K content was observed by the effect of potassium silicate at a concentration of 2 and 4 ppm by 8.08 and 21.2%, respectively, under 70% FC. However, the amount of decrease was 8.7 and 18.2%, respectively, under 60% FC (Fig. 1 a). The Si concentration was significantly improved by all potassium silicate applications under the water deficit conditions. The Si concentration increased in response to 4 mM potassium silicate under 80 and 60% FC (by 926 and 998%, respectively) compared to control plants (Fig. 1 b).

The results showed that the interaction of potassium silicate and drought stress significantly (P<0.01) affected electrolyte leakage, RWC, antioxidant activity, and MDA accumulation. Potassium silicate signifi-



Fig. 1 - Effect of exogenous application of potassium silicate on K (a), and Si (b) of daisy under water-stress conditions. Bars with a different letter differ significantly (P<0.05) according to the LSD test.

cantly inhibited the decrease in electrolyte leakage, whereas a more significant level of decrease was observed in the control plants at 60% FC. As shown in figure 2a, electrolyte leakage was increased by 102, 150, and 220% at 60% FC in control plants, 2 and 4 mM compared to 80% FC, respectively. RWC was reduced by 8.6 and 11.7% under drought stress (60% FC) in response to 2 and 4 mM potassium silicate compared to 80% FC. Compared with the control, the application of 4 mM potassium silicate significantly increased the RWC of plants, whereas 2 mM potassium silicate had no significant effect on the RWC compared to the control plants (Fig. 2 b).

The antioxidant activity increased significantly in response to drought stress, but the application of 2 and 4 mM potassium silicate under severe drought stress resulted in even higher values of antioxidant activity. Nonetheless, no significant difference was observed between potassium silicate-treated plants and the control plants at 80% FC. The antioxidant activity reached maximum values, increasing by 33.7 and 36.5% when the daisies were treated with 2 and

4 mM potassium silicate at 60% FC compared to the control plants, respectively (Fig. 2 c). The role of potassium silicate at 2 and 4 ppm was effective in reducing MDA accumulation under drought stress. Plants treated with potassium silicate under drought stress showed lower MDA levels in the following order: 2 and 4 mM of potassium silicate than control



Fig. 2 - Effect of exogenous application of potassium silicate on electrolyte leakage (a), relative water content (b), antioxidant activity (c), and MDA (d) of daisy under waterstress conditions. Bars with a different letter differ significantly (P<0.05) according to the LSD test.</p> plants. Thus, decreasing effect on this trait resulted from using 4 mM potassium silicate at 70 and 60% FC. The MDA peaked when the daisies were treated with distilled water and 2 mM potassium silicate at 60% FC. However, no significant difference was observed between the control plants and either of the 2 and 4 mM potassium silicate treatments under the effect of 80% FC (Fig. 2 d).

Drought stress significantly (P<0.05) increased the activities of antioxidant enzymes, catalase, peroxidase, superoxide dismutase, and aspartate peroxidase by 7.76-31.54%, 313-323%, 127-181%, and 4.22-99.35%, respectively, under drought stress (Fig. 3). In general, potassium silicate increased all of the mentioned enzyme activities, but this increase was higher in reponse to the 4 mM treatment under drought stress. The application of potassium silicate at 2 and 4 mM significantly improved the activity of CAT by 29.4 and 35.2%, respectively, in daisies grown at 60% FC compared to the control plants (Fig. 3 a). All potassium silicate treatments significantly improved the activity of POD under drought stress conditions. The POD activity increased in response to the 70% FC compared to the control, but decreased more at 60% FC, compared to 70% FC. As shown in figure 3 b, the activity of POD increased by 42.7 and 50.3% in plants treated with 2 and 4 mM potassium silicate, respectively, compared to the control plants under 60% FC.

Under the conditions of drought stress, the application of potassium silicate significantly increased the activity of APX. In response to 60% FC, the plants showed the highest APX activity. Although no significant differences were observed between potassium silicate-treated and control plants at 60% FC, a sharp increase in APX activity was observed when 4 mM potassium silicate was used along with drought stress. The application of 4 mM potassium silicate increased the activities of APX by 14.5% at 80% FC and by 87.8% at 60% FC compared to the control plants (Figs. 3c). The application of potassium silicate increased the SOD activity under drought stress conditions. This pattern of increase was more prominent (37.5 and 30.2%) at both potassium silicate levels along with moderate drought stress, compared to 80% FC, whereas it was least prominent in severe conditions (60% FC). The activity of SOD in daisy leaves increased by 28.4 and 21.5% at 60% FC, using potassium silicate at 2 and 4 mM, respectively, compared to well-watered plants (Fig. 3 d).

The data revealed that the chlorophyll (chl) a, b, and total chlorophyll contents were significantly

(P<0.05) affected by fertilizer and drought stress. Regarding chl a, b, and total chl in the leaves under drought stress, these parameters decreased in response to the drought stress severity. This downward trend was 34.68 and 55.4% higher in the case of Chl a, but was 2 and 6.25% lower in the case of Chl b when severity of drought increased. The application



Fig. 3 - Effect of exogenous application of potassium silicate on CAT (a), POD (b), APX (c), and SOD (d) of daisy under water-stress conditions. Bars with a different letter differ significantly (P<0.05) according to the LSD test.</p>

of potassium silicate at a concentration of 2 mM increased the chl a and chl b by 68 and 67%, respectively, at 70% FC. Furthermore, the mentioned values were increased by 41.4 and 97%, respectively, at 60% FC compared to the control. The highest content of chl a and b were observed when plants were treated with 4 mM potassium silicate at 80 and 70% FC (Figs. 4 a, b). During drought stress, total chl gradually decreased in response to greater intensity of drought stress. The total chl value decreased by 127% under 60% FC compared to 80% FC and by 98% compared to 70% FC. This value was also affected by potassium silicate under drought stress. Total Chl content was significantly improved by all potassium silicate appli-



Fig. 4 - Effect of exogenous application of potassium silicate on

test.

chlorohyll a (a), chlorohyll b (b), and total chlorohyll (c) of

daisy under water-stress conditions. Bars with a different

letter differ significantly (P<0.05) according to the LSD

cations under the drought stress conditions. Total chl

content increased by 7.4 and 72% in response to 4 mM potassium silicate at 80 and 70% FC, respectively (Fig. 4 c).

The application of potassium silicate significantly (P<0.05) increased the amount of carbohydrate and proline content in the leaves under drought stress. At 80% FC, potassium silicate-treated plants had no significant difference from the control plants in terms of carbohydrate content. By applying drought stress, an increase in carbohydrates was observed in the leaves when the plants were treated with potassium silicate. In response to 2 and 4 mM potassium silicate, the carbohydrate content increased by 31.3% and 26.6% at 70% FC, and by 60.8% and 53.2% at 60% FC, respectively, compared to the 80% FC (Fig. 5a). Regarding proline changes in the leaves, there was an increase of 4.7- and 1.6-fold by the effect of 60% FC and 70% FC, respectively, compared to the 80% FC. The application of 2 and 4 mM potassium silicate increased this parameter further by 75 and 42% at 70% FC, respectively, but by 385 and 312% at 60% FC, compared to the control plants. The highest proline



Fig. 5 - Effect of exogenous application of potassium silicate on carbohydrate (a), and proline (b) of daisy under waterstress conditions. Bars with a different letter differ significantly (P<0.05) according to the LSD test.

content was observed in drought-stressed plants (60% FC) treated with potassium silicate. However, no significant difference was observed between 2 and 4 mM potassium silicate under severe drought stress (Fig. 5b).

In general, potassium silicate (2 and 4 mM) at 80% FC significantly (P<0.01) increased the flavonoid content by 36 and 45.5% compared to the control plants. In the control plants. Increasing the severity of drought stress at 70 and 60% FC decreased this trait by 50.2 and 97.2%, respectively. The application of 2 and 4 mM potassium silicate increased the flavonoid content by 10.3 and 41.9%, respectively, at moderate water deficit (70% FC), but by 51.7 and 62.3%at severe water deficit (60% FC), respectively, compared to the 80% FC (Fig. 6 a). Figure 5b shows that the phenolic content increased in response to the intensity of drought stress, and the highest value was found at 60% FC. Total phenolic content increased significantly by 31.1 and 43.8% when potassium silicate-treated plants (4 mM) were under the effect of 80 and 70% FC, respectively, compared to the control plants. Potassium silicate (2 mM) increased the phenolic content by 27 and 46%, respectively. At 4 mM, it caused an increase of 39 and 57%, respectively, under the effect of 70 and 60% FC, compared to 80% FC. The highest anthocyanin content occurred when the potassium silicate-treated plants were grown at 80% FC. As for anthocyanin changes in the flowers, there was a decrease in response to the intensity of drought, but potassium silicate increased the value further. The anthocyanin content decreased in response to 4 mM potassium silicate at 60% FC, compared to 80% FC (11.2%). Both concentrations of potassium silicate significantly increased the anthocyanin content in the flowers by 228% at most, under the effect of 80% FC, compared to the control plants (Fig. 6 c).

4. Discussion and Conclusions

The application of potassium silicate improved nutrient uptake under drought stress. Si is not an essential nutrient but protects plants from a variety of biotic and abiotic stresses (Ranjan *et al.*, 2021). The highest concentration of K and Si in the plant occurred in response to 4 mM potassium silicate at 80% FC. These results emanate from the ability of plants to enhance root growth after potassium and silicon application, thereby increasing nutrient



Fig. 6 - Effect of exogenous application of potassium silicate on phenol (a), flavonoid (b), and anthocyanin (c) of daisy under water-stress conditions. Bars with a different letter differ significantly (P<0.05) according to the LSD test.</p>

uptake (Sustr *et al.*, 2019). The increase in K uptake is usually concomitant with a decrease in plasma membrane permeability and an increase in Si-induced H-ATP activity in plasma membranes (Zhu and Gong, 2014). K deficiency decreases the uptake and transfer of some nutrients by inhibiting enzymatic activities such as synthases, transferases, and kinases (Liu *et al.*, 2013). Si leads to more significant root activity, consequently, a greater amount of nutrient uptake. Sarto *et al.* (2014) attributed the beneficial effects of Si to its concentration in the leaves and stems of wheat. Si is also known to influence the development of apoplastic barriers in roots by controlling apoplastic pathways, followed by its translocation through the root apoplast to the shoot (Vaculík *et al.*, 2012). One explanation for the increased tolerance in plants growing under water-deficit conditions could be a decrease in transpiration through the stomata and cuticle due to Si application. Silicon not only affects nutrient availability and uptake but also nutrient translocation from the roots to the shoots (Greger *et al.*, 2018). In this study, drought significantly decreased the concentration (%) of K and Si. Also, drought stress can have a significant impact on plant nutrient ratios. Several studies have shown that drought can reduce nutrient uptake from the soil (Ge *et al.*, 2012; Bista *et al.*, 2018).

Under water-deficit conditions, potassium silicate reduces electrolyte leakage. Potassium silicate plays an important role in plant resistance to environmental stress. Within the plant, silicate is an immobile element that becomes a polymer gel and reduces the loss of ions from biomembranes after being deposited in the cell. The results of this study are consistent with a previous study by Othmani et al. (2021) who found that the more significant stability of the cell membrane in the presence of silicon was due to the hardening and strength of the cell wall. The water content of leaves under drought stress (70 and 60% FC) decreased compared to the 80% FC. Drought stress reportedly caused a decrease in leaf water content in most plants (Santos et al., 2021). In our study, the administration of potassium silicate led to an increase in the relative water content of leaves. A lower rate of water loss in silicate-potassium-fed plants can also be attributed to the lower transpiration of the plants. The accumulation of silicate in the lower epidermal cells reduces water loss through the cuticle. Si is deposited in plant tissues in the apoplast of the cell wall to form silica, thereby maintaining tissue integrity (Guerriero et al., 2016). In addition, potassium is primarily an important osmotic regulator in plants. Between 30 and 50% of the osmotic potential of leaf tissue is regulated by K (Turcios et al., 2021).

In the current study, potassium silicate reduced plant injury by decreasing MDA and increasing antioxidant activity. Malondialdehyde is the peroxidation product of unsaturated fatty acids in phospholipids. Therefore, the production of malondialdehyde under stress conditions can be used as a marker of lipid peroxidation (Ayala *et al.*, 2014). As a result of drought, the peroxidation of glycopeptides occurred in chloroplast thylakoid, followed by the formation of diacylglycerol, triacylglycerol, and free fatty acids, leading to an increase in malondialdehyde in plant tissues (Sofo et al., 2004). Fatty acids and lipids are reportedly sensitive to oxygen species and are rapidly oxidized. The results are in line with previous studies indicating a positive effect of potassium silicate on malondialdehyde levels in damask rose (Rosa damascena Miller) under drought stress (Farahani et al., 2020). The ability of plants to scavenge free radicals was impaired by both drought and the addition of potassium silicate compared to the control plants. DPPH inhibition levels were below 70% and 60% FC at 4 mM potassium silicate compared to the control. Under drought stress, DPPH levels increased, and potassium silicate further enhanced the DPPH levels (Zahedi et al., 2020).

In this study, the administration of potassium silicate under drought stress conditions (i.e. the application of irrigation water to maintain 70 and 60% FC) increased all antioxidant enzyme activities. An increase in antioxidant activity in the leaves occurred in response to both potassium silicate concentrations and drought stress. The high activity of antioxidant enzymes such as CAT, POD, APX, and SOD in plants is an adaptive mechanism that protects cells from oxidative damage by reducing the concentration of hydrogen peroxide generated by cellular metabolism (Jan et al., 2022). Improving potassium concentration leads to an increase in photosynthetic products, the control of ionic balance, osmotic regulation, and an increase in enzymatic activity. By stimulating the activity of POD and APX through the detoxification of hydrogen peroxide, Si prevents oxidative stress and inhibits the production of hydroxyl radicals (Kim et al., 2017). In agreement with the current results, Ahmad et al. (2019) reported that using silica on mung beans (Vigna radiata L.) increased catalase and superoxide dismutase activities under drought stress. Superoxide dismutase is an enzyme that converts superoxide free radicals into hydrogen peroxide and oxygen while playing an important role in protecting cells from the negative effects of free radicals. SOD is the first line of defense of cells against free radicals under stress conditions (Ighodaro et al., 2018). The effects of Si nutrition on SOD activity and free radical elimination have been reported in the available literature (Geng et al., 2018). Gong et al. (2005) reported that using potassium silicate increased the activity of antioxidant enzymes in wheat (Triticum aestivum L.) under drought stress. The removal of reactive oxygen species decreases cell membrane permeability and

increases the activity of catalase, peroxidase, and superoxide dismutase, which indirectly decrease cell membrane lipid peroxidation and reduce the amount of malondialdehyde (Sharma *et al.*, 2012).

The application of potassium silicate along with drought stress increased the amounts of photosynthetic pigments. A decrease in chlorophyll content occurred due to drought stress and was accompanied by an increase in the production of oxygen radicals in the cells. The radicals usually cause peroxidation, and consequently, the degradation of photosynthetic pigments. The effect of potassium silicate on the stability of plant pigments usually results from the accumulation of silicate in the epidermal cells, which has an indirect protective effect on the photosynthetic establishments, thereby reducing the stress-induced damage to photosynthetic pigments. Similar to the current results, a moderating effect of potassium silicate was reportedly observed on the chlorophyll content of Rosmarinus officinalis L. plants (Waly et al., 2019).

The results showed that the concentration of osmotic regulators (i.e. proline and total carbohydrates) increased significantly when potassium silicate was applied under water-deficit conditions. Silicon and potassium increase the production of carbohydrates and proline by increasing the osmotic potential, possibly through the accumulation of free radicals produced by the plant. They are thought to play an adaptive role in mediating osmotic adjustment and protecting subcellular structures in stressed plants (Hajiboland et al., 2017). These effects suggest that potassium and silicon may enhance leaf osmotic potential by converting starch to soluble sugars, especially under severe drought stress (Zahoor et al., 2017). It appears that potassium silicate stimulates carbohydrate production and, thus, alters the metabolism of plant-absorbed K and its conversion to proteins (Hafez et al., 2021). Si can directly or indirectly induce the biosynthesis of proline. Garg and Sing (2018) showed that the application of Si increased the activity of pyrroline-5-carboxylate synthetase (P₋CS) and glutamate dehydrogenase (GDH). In addition, the increase in proline because of potassium silicate treatment may highlight the importance of potassium and silicon in protecting cell membranes and maintaining relative water content under inadequate irrigation conditions. In this context, using silica on borage (Borago officinalis L.) plants reportedly increased the amount of proline in the leaves (Gagoonani et al., 2011). In agreement with these results, Ibrahim et al. (2020)

reported that potassium silicate increased the proline content of maize plants under drought stress.

By reducing vegetative growth and altering the anatomical structure of the plant through the induction of secondary stress, e.g. oxidative stress, the effect of drought stress usually cause changes in the pathways of synthesis that make secondary compounds and metabolites (Ahanger et al., 2017). Polyphenols can improve plant tolerance to drought stress and play an important role as a carbon sink at times of stress. These effects may explain significant improvements in total phenolics in daisies because of their exposure to drought stress (Fig. 6 a). The increase in total soluble phenols in response to the application of K₂SiO₃ under drought stress could be a supporting effect of Si, thereby increasing plant tolerance, especially under water-deficit conditions. Fouda et al. (2021) found that potassium silicate increased the total flavonoid content in field beans. Feeding plants with Si and K-containing compounds has reportedly resulted in changes in the expression pattern of many genes. In particular, feeding plants with potassium- and silicon-containing compounds has led to changes in genes that encode enzymes involved in the phenylpropanoid pathway (Wang et al., 2017). Indeed, the increase in phenylalanine ammonia-lyase activity is a common feature in plants treated with silicon, thereby enabling an increase in the synthesis of phenolic compounds. Potassium silicate increases polyphenols in many plants by activating enzymes that are relevant to the phenol production pathway, such as the phenylalanine ammonialyase (Vega et al., 2019).

The ability of plants to tolerate drought could be mainly explained by an increase in flavonoid content since flavonoids are compounds with strong antioxidant activity. Perin et al. (2019) suggested that the relationships between ABA metabolism, phenylpropanoid, flavonoid, and anthocyanin pathways can reduce drought stress. Probably, this could be one of the main reasons for the better tolerance of plants to drought stress. Drought largely affects the average performance of plant traits by reducing their properties, leading to a decrease in the associated anthocyanin content. Under drought stress, the anthocyanin content decreased, but potassium silicate increased the anthocyanin content of flowers (Fig. 6 c). These results are consistent with a previous study by Cirillo et al. (2021) in which anthocyanin content was reduced by the effects of stress. Jafari et al. (2015) reported that silicon treatment under osmotic stress significantly increased the amount of nonenzymatic antioxidants (e.g. anthocyanins, flavonoids, and total phenolic compounds) and nutrients (Si, K⁺, and Ca⁺²) in cucumber plants.

References

- ABEDI T., PAKNIYAT H., 2010 Antioxidant enzyme changes in response to drought stress in ten cultivars of oil seed rape (Brassica napus L.). - Czech J. Gen. Plant Breed., 46: 27-34.
- AHANGER M.A., TOMAR N.S., TITTAL M., ARGAL S., AGAR-WAL R.M., 2017 - Plant growth under water/salt stress: *ROS production, antioxidants and significance of added potassium under such conditions.* - Physiol. Mol. Biol. Plants, 23: 731-744.
- AHMAD P., AHANGER M.A., ALAM P., ALYEMENI M.N., WIJAYA L., ALI S., ASHRAF M., 2019 - Silicon (Si) supplementation alleviates NaCl toxicity in mung bean [Vigna radiata (L.) Wilczek] through the modifications of physio-biochemical attributes and key antioxidant enzymes. - J. Plant Growth Regul., 38: 70-82.
- AYALA A., MUÑOZ M.F., ARGÜELLES S., 2014 Lipid peroxidation: production, metabolism, and signaling mechanisms of malondialdehyde and 4-hydroxy-2-nonenal. -Oxidative Medicine Cellular Longevity, 2014: 360438.
- BATES L.S., WALDREN R.P., TEARE I.D., 1973 *Rapid determination of free proline for water-stress studies.* - Plant Soil, 39: 205-207.
- BISTA D., HECKATHORN S., JAYAWARDENA D., MISHRA S., BOLDT J., 2018 - Effects of drought on nutrient uptake and the levels of nutrient-uptake proteins in roots of drought-sensitive and -tolerant grasses. - Plants, 7(2): 28.
- BUKHARI M.A., AHMAD Z., ASHRAF M.Y., AFZAL M., NAWAZ F., NAFEES M., JATOI W.N., MALGHANI N.A., SHAH A.N., MANAN A., 2020 - *Silicon mitigates drought stress in wheat* (Triticum aestivum *L.*) *through improving photosynthetic pigments, biochemical and yield characters.* - Silicon, 2020: 1-16.
- CALANCA P.P., 2017 Effects of abiotic stress in crop production, pp. 165-180. - In: AHMED M., and C. STOCKLE (eds.) Quantification of climate variability, adaptation and mitigation for agricultural sustainability. Springer Verlag, Switzerland, pp. 437.
- CAMPBELL G.S., MULLA D.J., 1990 *Measurement of soil water content and potential*. - University of California Agricultural Experiment Station, Berkeley, CA.
- CIRILLO V., D'AMELIA V., ESPOSITO M., AMITRANO C., CARILLO P., CARPUTO D., MAGGIO A., 2021 -Anthocyanins are key regulators of drought stress tolerance in tobacco. - Biology, 10: 139.
- DACOSTA M., HUANG B., 2007 Change in antioxidant activities and lipid peroxidation for bent-grass species

in responses to drought stress. - J. Amer. Soc. Hort. Sci., 132: 319-326.

- DALLAGNOL L.J., RODRIGUES F.A., TANAKA F.A.O., AMOR-IM L., CAMARGO L.E.A., 2012 - *Effect of potassium silicate on epidemic components of powdery mildew on melon.* - Plant Pathol., 61: 323-330.
- FARAHANI H., SAJEDI N.A., MADANI H., CHANGIZI M., NAEINI M.R., 2020 - Physiological and biochemical responses of Damask rose (Rosa damascena Miller) to potassium silicate application under water deficit stress. - Notulae Botanicae Horti Agrobotanici Cluj-Napoca, 48(3): 1560-1572.
- FOUDA S.E., EL-SAADONY F.M., SAAD A.M., SAYED S.M., EL-SHARNOUBY M., EL-TAHAN A.M., EL-SAADONY M.T. 2021 - Improving growth and productivity of faba bean (Vicia faba L.) using chitosan, tryptophan, and potassium silicate anti-transpirants under different irrigation regimes. - Saudi J. Biolog. Sci., 29(2): 955-962.
- GAGOONANI S., ENTESHARI S., DELAVAR K., MOOHAMAD-BAGHER B., 2011 - Interactive effects of silicon and aluminum on the malondialdehyde (MDA), proline, protein and phenolic compounds in Borago officinalis L. - J. Med. Plants Res., 5: 5818-5827.
- GAO S., WANG Y., YU S., HUANG Y., LIU H., CHEN W., HE X., 2020 - Effects of drought stress on growth, physiology and secondary metabolites of Two Adonis species in Northeast China. - Scientia Hortic., 259: 108795.
- GARG N., SINGH S., 2018 Arbuscular mycorrhiza Rhizophagus irregularis and silicon modulate growth, proline biosynthesis and yield in Cajanus cajan L. Millsp. (pigeonpea) genotypes under cadmium and zinc stress.
 - J. Plant Growth Regul., 37: 46-63.
- GE T.D., SUN N.B., BAI L.P., TONG C.L., SUI F.G., 2012 -Effects of drought stress on phosphorus and potassium uptake dynamics in summer maize (Zea mays) throughout the growth cycle. - Acta Physiologiae Plantarum, 34: 2179-2186.
- GENG A., WANG X., WU L., WANG F., WU Z., YANG H., CHEN Y., WEN D., LIU X., 2018 - Silicon improves growth and alleviates oxidative stress in rice seedlings (Oryza sativa L.) by strengthening antioxidant defense and enhancing protein metabolism under arsanilic acid exposure. - Ecotoxicology Environ. Safety, 158: 266-273.
- GONG H., ZHU X., CHEN K., WANG S., ZHANG C., 2005 -Silicon alleviates oxidative damage of wheat plants in pots under drought. - Plant Sci., 169: 313-321.
- GREGER M., LANDBERG T., VACULÍK M., 2018 Silicon influences soil availability and accumulation of mineral nutrients in various plant species. - Plants, 7: 41.
- GUAN G., WANG Y., CHENG H., JIANG Z., FEI J., 2015 -Physiological and biochemical response to drought stress in the leaves of Aegiceras corniculatum and Kandelia obovata. - Ecotoxicology, 24: 1668-1676.
- GUERRIERO G., HAUSMAN J.F., LEGAY S., 2016 Silicon and the plant extracellular matrix. Frontiers in Plant Sci.,

7: 463.

- GUSTA L.V., WISNIEWSKI M., NESBITT N.T., TANINO K.T., 2003 - Factors to consider in artificial freeze tests. -Acta Horticulturae, 618: 493-507.
- HAFEZ E.M., OSMAN H.S., EL-RAZEK U.A.A., ELBAGORY M., OMARA A.E.D., EID M.A., GOWAYED S.M., 2021 -Foliar-applied potassium silicate coupled with plant growth-promoting rhizobacteria improves growth, physiology, nutrient uptake and productivity of faba bean (Vicia faba L.) irrigated with saline water in saltaffected soil. - Plants, 10: 894.
- HAJIBOLAND R., CHERAGHVAREH L., POSCHENRIEDER C., 2017 - Improvement of drought tolerance in tobacco (Nicotiana rustica L.) plants by silicon. - J. Plant Nutr., 40: 1661-1676.
- HAWORTH M., MARINO G., COSENTINO S.L., BRUNETTI C., DE CARLO A., AVOLA G., RIGGI E., LORETO F., CENTRIT-TO M., 2018 - Increased free abscisic acid during drought enhances stomatal sensitivity and modifies stomatal behaviour in fast growing giant reed (Arundo donax L.). - Environ. Exper. Bot., 147: 116-124.
- IBRAHIM M.F.M., ABD EL-SAMAD G., ASHOUR H., EL-SAWY A.M., HIKAL M., ELKELISH A., ABD EL-GAWAD H., EL-YAZIED A.A., HOZZEIN W.N., FARAG R., 2020 -Regulation of agronomic traits, nutrient uptake, osmolytes and antioxidants of maize as influenced by exogenous potassium silicate under deficit irrigation and semiarid conditions. - Agronomy, 10(8): 1212.
- IGHODARO O.M., AKINLOYE O.A., 2018 First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. -Alexandria J. Med., 54: 287-293.
- JAFARI S.R., ARVIN S.M.J., KALANTARI K.M., 2015 -Response of cucumber (Cucumis sativus L.) seedlings to exogenous silicon and salicylic acid under osmotic stress. - Acta Biologica Szegediensis, 59: 25-33.
- JAISWAL P.C., 2003 Soil, plant and water analysis. -Kalyani Publishers, Ludhiana, New Delhi, India, pp. 450.
- JAN A.U., HADI F., DITTA A., SULEMAN M., ULLAH M., 2022 - Zinc-induced anti-oxidative defense and osmotic adjustments to enhance drought stress tolerance in sunflower (Helianthus annuus L.). - Environ. Exper. Bot., 193: 104682.
- KEDARE S.B., SINGH R.P., 2011 Genesis and development of DPPH method of antioxidant assay. - J. Food Sci. Technol., 48: 412-422.
- KIM Y.H., KHAN A.L., WAQAS M., LEE I.J., 2017 Silicon regulates antioxidant activities of crop plants under abiotic-induced oxidative stress: a review. - Frontiers Plant Sci., 8: 510.
- KUSVURAN S., DASGAN H.Y., 2017 Effects of drought stress on physiological and biochemical changes in Phaseolus vulgaris L. Legume Research, 40(1): 55-62.
- LIU C.H., CHAO Y.Y., KAO C.H., 2013 Effect of potassium deficiency on antioxidant status and cadmium toxicity

in rice seedlings. - Botanical Studies, 54: 1-10.

- MCCREADY R.M., GUGGOLZ J., SILVIERA V., OWENS H.S., 1950 - Determination of starch and amylose in vegetables. - Analytical Chem., 22: 1156-1158.
- MOUSSA S., SHAMA M., 2019 Mitigation the adverse effects of irrigation water salinity on potato crop using potassium silicate foliar application. - Middle East J. Appl. Sci., 9: 804-819.
- NAGATA M., YAMASHITA I., 1992 Simple method for simultaneous determination of chlorophyll and carotenoids in tomato fruit. - Nippon Shokuhin Kogyo Gakkaish, 39: 925-928.
- OTHMANI A., AYED S., BEZZIN O., FAROOQ M., AYED-SLAMA O., SLIM AMARA H., YOUNES M.B., 2021 - Effect of silicon supply methods on durum wheat (Triticum durum Desf.) response to drought stress. - Silicon, 13: 3047-3057.
- PERIN E.C., DA SILVA MESSIAS R., BOROWSKI J.M., CRIZEL R.L., SCHOT I.B., CARVALHO I.R., ROMBALDI C.V., GALLI V., 2019 - ABA-dependent salt and drought stress improve strawberry fruit quality. - Food Chem., 271: 516-526.
- PIECZYNSKI M., MARCZEWSKI W., HENNIG J., DOLATA J., BIELEWICZ D., PIONTEK P., WYRZYKOWSKA A., KRUSIEWICZ D., STRZELCZYK-ZYTA D., KONOPKA-POS-TUPOLSKA D., KRZESLOWSKA M., JARMOLOWSKI A., SZWEYKOWSKA KULINSKA Z., 2013 - Down-regulation of CBP80 gene expression as a strategy to engineer a drought-tolerant potato. - Plant Biotech. J., 11: 459-469.
- PRĂVĂLIE R., 2016 Drylands extent and environmental issues. A global approach. Earth-Science Reviews, 161: 259-278.
- QI J., SUN S., YANG L., LI M., MA F., ZOU Y., 2019 -Potassium uptake and transport in apple roots under drought stress. - Hortic. Plant J., 5: 10-16.
- RANJAN A., SINHA R., BALA M., PAREEK A., SINGLA-PAREEK S.L., SINGH A.K., 2021 - Silicon-mediated abiotic and biotic stress mitigation in plants: Underlying mechanisms and potential for stress resilient agriculture. -Plant Physiol. Bioch., 163: 15-25.
- RAZA A., ASHRAF F., ZOU X., ZHANG X., TOSIF H., 2020 -Plant adaptation and tolerance to environmental stresses: mechanisms and perspectives, pp. 117-145. -In: HASANUZZAMAN M. (ed.) Plant ecophysiology and adaptation under climate change: mechanisms and perspectives I. General consequences and plant responses. Springer, Singapore, pp. 859.
- REYNOLDS O.L., KEEPING M.G., MEYER J.H. 2009 Siliconaugmented resistance of plants to herbivorous insects: A review. - Ann. Appl. Biol., 155: 171-186.
- ROMERO-ARANDA M.R., JURADO O., CUARTERO J., 2006 -Silicon alleviates the deleterious salt effect on tomato plant growth by improving plant water status. - J. Plant Physiol., 163: 847-855.

SAIRAM R.K., RAO K.V., SRIVASTAVA G.C., 2002 -

Differential response of wheat genotypes to long term salinity stress in relation to oxidative stress, antioxidant activity and osmolyte concentration. - Plant Sci., 163: 1037-1046.

- SANTOS M., BARROS V., LIMA L., FROSI G., SANTOS M.G., 2021 - Whole plant water status and non-structural carbohydrates under progressive drought in a Caatinga deciduous woody species. - Trees, 35: 1257-1266.
- SARTO M.V.I.M., DO CARMO LANA M., RAMPIM L., ROSSET J.S.E., WOBETO J.R., ECCO M., BASSEGIO D., DA COSTA P.F., 2014 - Effect of silicate on nutrition and yield of wheat. - African J. Agric. Res., 9: 956-962.
- SHARMA P., JHA A.B., DUBEY R.S., PESSARAKLI M., 2012 -Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. - J. Bot., 2012: 1-26.
- SIATKA T., KAŠPAROVÁ M., 2010 Seasonal variation in total phenolic and flavonoid contents and DPPH scavenging activity of Bellis perennis L. flowers. -Molecules, 15: 9450-9461.
- SINGLETON V., ROSSI J.A., 1965 Colorimetry of total phenolics with phosphomolybdic-phosphotungstic acid reagents. - Amer. J. Enol. Vitic., 16: 144-158.
- SOFO A., DICHIO B., XILOYANNIS C., MASIA A., 2004 -Lipoxygenase activity and proline accumulation in leaves and roots of olive trees in response to drought stress. - Physiologia Plantarum, 121: 58-65.
- SUKWATTANASINIT T., BURANA-OSOT J., SOTANAPHUN U., 2007 - Spectrophotometric method for quantitative determination of total anthocyanins and quality characteristics of roselle (Hibiscus sabdariffa). - Planta Medica, 73: 1517-1522.
- SUSTR M., SOUKUP A., TYLOVA E., 2019 Potassium in root growth and development. Plants, 8: 435.
- TURCIOS A.E., PAPENBROCK J., TRÄNKNER M., 2021 -Potassium, an important element to improve water use efficiency and growth parameters in quinoa (Chenopodium quinoa) under saline conditions. - J. Agron. Crop Sci., 207:,1-13.
- VACULÍK M., LANDBERG T., GREGER M., LUXOVÁ M., STOLÁRIKOVÁ M., LUX A., 2012 - Silicon modifies root anatomy, and uptake and subcellular distribution of cadmium in young maize plants. - Annals Bot., 110: 433-443.

- VEGA I., NIKOLIC M., PONTIGO S., GODOY K., MORA M.D.L.L., CARTES P., 2019 - Silicon improves the production of high antioxidant or structural phenolic compounds in barley cultivars under aluminum stress. -Agronomy, 9: 388.
- VELIKOVA V., YORDANCV I., EDREVA A., 2000 Oxidative stress and some antioxidant systems in acid rain-treated bean plants. Protective role of exogenous polyamines. - Plant Sci., 151: 59-66.
- WALY A.A., EL-FATTAH A., HASSAN M.A.E., EL-GHADBAN E.A.E., ABD ALLA A.S., 2019 - Effect of foliar spraying with seaweeds extract, chitosan and potassium silicate on Rosmarinus officinalis L. plants in sandy soil. -Scientific J. Flowers Ornam. Plants, 6: 191-209.
- WANG M., GAO L., DONG S., SUN Y., SHEN Q., GUO S., 2017 - *Role of silicon on plant-pathogen interactions*. -Front. Plant Sci., 8: 701.
- WANG M., WANG R., MUR L.A.J., RUAN J., SHEN Q., GUO S., 2021 Functions of silicon in plant drought stress responses. Hortic. Res., 8: 254.
- WEERAHEWA H.L.D., DAVID D., 2015 Effect of silicon and potassium on tomato anthracnose and on the postharvest quality of tomato fruit (Lycopersicon esculentum Mill.). - J. Nat. Sci. Foundation of Sri Lanka, 43: 273-280.
- ZAHEDI S.M., MOHARRAMI F., SARIKHANI S., PADERVAND M., 2020 - Selenium and silica nanostructure-based recovery of strawberry plants subjected to drought stress. - Scientific Reports, 10: 1-18.
- ZAHOOR R., ZHAO W., ABID M., DONG H., ZHOU Z., 2017 -Potassium application regulates nitrogen metabolism and osmotic adjustment in cotton (Gossypium hirsutum L.) functional leaf under drought stress. - J. Plant Physiol., 215: 30-38.
- ZHANG Y., SHI Y., GONG H., ZHAO H.L., LI H.L., HU Y.H., WANG Y.C., 2018 - Beneficial effects of silicon on photosynthesis of tomato seedlings under water stress. - J. Integrative Agric., 17: 2151-2159.
- ZHU Y., GONG H., 2014 Beneficial effects of silicon on salt and drought tolerance in plants. - Agronomy Sustainable Develop., 34: 455-472.
- ZOU Y., LU Y., WEI D., 2004 Antioxidant activity of *flavonoid-rich extract of* Hypericum perforatum *L*. in vitro. - J. Agric. Food Chem., 52: 5032-5039.